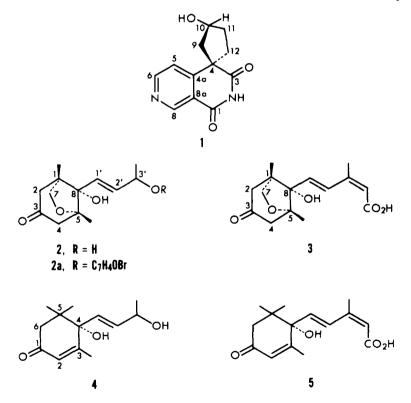
AN INVESTIGATION OF THE ANTITUMOR ACTIVITY OF SESBANIA DRUMMONDII

RICHARD G. POWELL and CECIL R. SMITH, JR.

Northern Regional Research Center, Agricultural Research, Science and Education Administration, U.S. Department of Agriculture,¹ Peoria, Illinois 61604

ABSTRACT.—Extracts of Sesbania drummondii (Rydb.) Cory seed are active in vivo in the P-388 lymphocytic leukemia system and cytotoxic in the KB assay. Systematic fractionation of S. drummondii has yielded sesbanine (1), a novel spirocyclic alkaloid based on the 2,7-naphthyridine nucleus, and drummondol (2), a new abscisic acid (5) metabolite. Sesbanine and drummondol are present in highly cytotoxic fractions; however, these purified compounds are essentially devoid of antitumor activity.

Sesbania drummondii (Rydb.) Cory (Fabaceae) is a perennial shrub or small tree inhabiting seasonally wet waste places and previously cultivated fields along the Coastal Plain from Florida to Texas. Seeds of S. drummondii, also known as coffeebean, rattlebrush, and rattlebox, are toxic to livestock, with the minimum lethal amount for sheep reported as about 0.1% of the animal's weight (1). In a previous report, we revealed that ethanolic extracts of the seed were cytotoxic



¹The mention of firm names or trade products does not imply that they are endorsed or recommended by the U.S. Department of Agriculture over other firms or similar products not mentioned.

in the KB cell culture and gave marked inhibition of P-388 lymphocytic leukemia (PS) in mice (2). Large-scale systematic fractionation of S. drummondii has revealed two new compounds, sesbanine (1) and drummondol (2). Although present in highly cytotoxic concentrates, 1 and 2 are essentially inactive after purification by hplc. We now report details of the isolation and characterization of these novel compounds.

An ethanol extract obtained from 454 kg of S. drummondii seed gave significant PS activity (T/C 175% at 45 mg/kg). Partition of the residue between aqueous ethanol and petroleum ether concentrated activity in the aqueous layer, and repeated washing of the aqueous layer with 1-butanol gave further concentration of activity in the butanol layer (T/C 153% at 10 mg/kg). Butanol solubles, 6.7 kg, were further fractionated by countercurrent distribution (ccd) using a three component biphasic system of water, ethyl acetate, and methanol in a 2:2:1 ratio. A preliminary 10-tube ccd experiment using 250 ml (125 ml each phase) of solvent per tube with 27 g of butanol solubles is summarized in table 1.

TABLE 1. Cytotoxic (KB) and antileukemic (PS) activity of fractions obtained by countercurrent distribution of *S. drummondii* butanol solubles.^a

CCD fraction and recovered weight (g)	KB cytotoxicity ED50, mcg/ml	$\begin{array}{l} Maximum \ PS \ activity \\ T/C \ (dose, \ mg/kg) \end{array}$	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	15.0 2.5 0.77 0.40 0.48 1.2 26.0 2.4 29.0 24.0	$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	

^aSolvent system was prepared by mixing water, ethyl acetate, and methanol in a 2:2:1 ratio.

Sugars and other hydrophilic compounds were prominent in ccd fractions 1 and 2, whereas fractions 7–10 were rich in lipophilic materials. Fractions 3–6 were highly cytotoxic (KB cell culture) and demonstrated *in vivo* PS activity at dose levels down to 0.5 mg/kg. These intermediate fractions proved to be rich in polyphenols including catechin, gallocatechin, and afzelechin. The scatter of high T/C values in all 10 ccd fractions indicates that several compounds of widely different polarity contribute to the observed PS activity. Scale-up of the ccd procedure gave comparable results for the remaining butanol solubles, and fractions 4 (455 g), 5 (453 g), and 6 (513 g) were selected for further study.

Removal of polyphenolic material was achieved by partition of ccd fractions between ethyl acetate and 5% aqueous sodium carbonate. Cytotoxic materials concentrated in the ethyl acetate layer. Further concentration of KB activity was achieved by chromatography on silica, chromatography on Sephadex G-10, and, finally, by hplc in two stages on reversed-phase columns. After 280-fold concentration, from ccd fractions 4-6 (1.4 kg) through chromatography on Sephadex (5.2 g), the cytotoxicity and PS activity of the most active fraction had not increased significantly. This observation suggests that the active principles of S. drummondii are relatively labile and that extensive degradation of the active components occurred during these isolation steps.

Sesbanine (1) was isolated, $1.1 \ge 10^{-5}\%$ yield, as one of a series of overlapping peaks from a C₁₈ bonded column with water-methanol (9:1) as the eluting solvent. Crude 1 was cytotoxic, $ED_{50} = 4.7 \ge 10^{-2} \text{ mcg/ml}$. However, recrystallization from methanol gave 1, mp 240–243°, which was only marginally cytotoxic and was inactive against PS *in vivo* at levels up to 1.4 mg/kg. The structure of 1 was revealed in an earlier x-ray crystallographic study (3) and has been confirmed by total synthesis (4, 5).

Drummondol (2) was isolated as a colorless oil, $3.2 \times 10^{-5}\%$ yield, with retention time approximately $\frac{3}{4}$ that of 1 on a C₁₈ column under similar chromatographic conditions. Compound 2 was inactive against KB; ED₅₀=5.4 x 10¹ mcg/ml. The structure of 2 was determined by detailed analysis of the ¹H and ¹³C spectral data (table 2). Acylation of 2 with *p*-bromobenzoyl chloride gave 2a, which

¹ H assignment	Chemical shift ^b	¹³ C assignment	Chemical shift ^b
C-1 methyl C-2 HA. HB. C-4 CH ₂ . C-5 methyl. C-7 HA. HB. C-1' vinyl H. C-2' vinyl H. C-3' H. C-3' methyl.	2.60 (dd); $J=18,2$ 2.35 (d); $J=18$ 2.61 (s) 1.20 (s) 3.88 (dd); $J=8,2$ 3.72 (d); $J=8$ 5.86 (q); $J=15,1$ 6.20 (q); $J=15,6$	C-1 methyl C-2 C-3 C-4 C-5 C-5 methyl C-7 C-8 C-1' C-2' C-3' C-3' methyl	$\begin{array}{c} 47.7 \ (s) \\ 15.4 \ (q) \\ 52.6 \ (t)^{\circ} \\ 206.9 \ (s) \\ 53.1 \ (t)^{\circ} \\ 81.9 \ (s) \\ 18.9 \ (q) \\ 77.3 \ (t) \\ 85.7 \ (s) \\ 124.1 \ (d)^{d} \\ 140.0 \ (d)^{d} \\ 68.0 \ (d) \\ 24.2 \ (q) \end{array}$

TABLE 2. ¹H and ¹³C nmr assignments for drummondol (2).^a

^aOur numbering of 2 is that of a 6-oxabicyclo[3,2,1]-octane and differs from that of previous work (6, 8) which attempted to demonstrate the close relationship of 3 and 5.

^bChemical shifts in ppm (δ) and coupling constants (J) as Hz (cps). Extensive decoupling was used to verify assignments.

•Assignments may be reversed.

^dAssignments may be reversed.

exhibited a downfield shift of the C-3' proton signal from \$4.45 to \$5.58. The 'H spectrum of 2 clearly shows that there is long range coupling (2 Hz) between the downfield C-7 oxymethylene signal (\$3.88) and the C-2 methylene signal centered at \$2.60. Other appropriate signals compare well with the 'H spectrum of the methyl ester of phaseic acid (3) (6). The ms of 2 and 2a were similar in that the highest ion observed was m/e 222 for both. This corresponded to M⁺-H₂O for 2 and M⁺-C₇H₅O₂Br for 2a.

It seems probable that 2 arises from 3 by oxidative removal of two carbons in a manner similar to that proposed for the formation of vomifoliol (4) from abscisic acid (5) (7). Alternatively, 2 could arise from 4 by an oxidative pathway similar to that demonstrated for the conversion of 5 to 3 (8).

We are continuing our investigations of fractions derived from S. drummondii seed extracts in an effort to establish the true antileukemic principles.

EXPERIMENTAL²

PLANT MATERIAL.—Seed of Sesbania drummondii (Rydb.) Cory was harvested from the wild in Texas during November 1975 (PR-44884). The collection was arranged and authenticated by Dr. Robert E. Perdue of the Medicinal Plant Resources Laboratory, USDA, Beltsville, Maryland.

EXTRACTION AND FRACTIONATION.—Seed material (454 kg) was ground in a Fitz mill to a particle size of 0.5-2.0 mm in diameter immediately prior to extraction. The ground material was placed in a 2270 liter tank containing 908 liters of 95% ethanol, heated to 50°, and stirred overnight. The ethanol extract was removed and the seed material was similarly extracted four more times, each with 450 liters of 95% ethanol. Ethanol extracts were combined and concentrated in a vacuum evaporator to a light syrup estimated to contain 25 kg of alcohol solubles. The concentrate was diluted with sufficient water to give 227 liters of an ethanol-water (1:3) solution, which was defatted by partitioning three times with a total of 400 liters of petroleum ether. Petroleum ether solubles amounted to 10.1 kg. Alcohol was then removed from the aqueous layer by concentrating to a volume of 45 liters; an equal volume of water was added, and the resulting solution was extracted once with 90 liters of 1-butanol and twice with 45 liters of 1-butanol. Butanol solubles were concentrated in an evaporator yielding 6.7 kg of a thick syrup.

Butanol soluble material, 6.6 kg, was then passed through a 10-carboy ccd in four batches. Each carboy contained 16 liters of a three-component biphasic solvent: water, ethyl acetate, and methanol in a 2:2:1 ratio, adjusted to equal phase volumes. Based on activity data of a preliminary small scale ccd (table 1) and similar data on the large scale ccd, fractions 4 (455 g), 5 (453 g), and 6 (513 g) were selected for further study. Each of the three ccd fractions was divided into two portions. To further concentrate activity of the six fractions thus obtained, each was dissolved in 9 liters of 5% Na₂CO₃ solution and extracted four times with 3 liters of ethyl acetate. In this manner, considerable polar and acidic (phenolic) material was removed and 238 g of ethyl acetate solubles remained. This material was divided into 15 portions ranging from 10-25 g: each portion was chromatographed on 250 g of silica in a 6 cm ID column. Each column was prepared in chloroform and eluted with a gradient consisting of 1.5 liters chloroform-ethyl acetate (1:1), 1.5 liters ethyl acetate, 1.5 liters ethyl acetatemethanol (1:1), and 0.75 liter of methanol. Forty 100-ml fractions collected from each run were combined into four, based on cytotoxicity the tle similarity, and similar fractions from all 15 runs were combined (48 g). Tle of this material on silica plates, developed with chloroform-methanol-water (40:10:1), gave many spots in the R 0.45-0.76 range.

The fractions of intermediate polarity, from chromatography on silica (48 g), were rechromatographed on a 3.5 cm ID column packed with Sephadex G-10 in 11 portions of 2-6 g each. The columns, prepared in water, were eluted in sequence with 1 liter of water, 500 ml of water-ethanol (1:1), and 500 ml of 95% ethanol. The first 1 liter fraction from each column was found to contain all of the cytotoxic material recovered and, when combined, it amounted to 5.2 g: $ED_{50}=3.9 \times 10^{-1} \mu g/ml$ (KB) and T/C=174 at 20 mg/kg (PS). The active concentrate from chromatography on Sephadex was then subjected to reversed-

The active concentrate from chromatography on Sephadex was then subjected to reversedphase liquid chromatography on a 2 m x 1 cm column packed with C_{15} Porasil B with watermethanol (4:1) as solvent at a flow rate of 9 ml/min. The most active fraction eluted with a retention time of 22-23 min and was resolved into a series of 16 peaks by further hple on a $C_{15} \mu$ Bondapak column (30 cm x 7.9 mm) with the same solvent and a flow rate of 2 ml/min. Drummondol (2) was obtained as a sharp peak with retention time of 17.5 min, and sesbanine (1) was a major component of the most active fraction having a broad peak with retention time of 20-25 min. Further chromatography on the same column with water-methanol (9:1) gave crude 1, retention time 65-73 min, $ED_{50}=4.7 \times 10^{-2} \mu g/ml$.

²Melting points were determined on a Fisher-Johns block and are uncorrected. Optical rotations were determined with a Cary Model 60 recording spectropolarimeter and ir analyses were done on 1% solutions in CHCl₃ with a Perkin-Elmer model 700 instrument. Low-resolution mass spectra were obtained on a DuPont (CEC) 21-492-1 spectrometer, and high resolution data with a Kratos MS-30 instrument. ¹H nmr spectra were measured with a Varian XL-100 spectrometer and ¹³C spectra were measured on a Bruker WH-90 Fourier transform spectrometer in CDCl₃ solutions unless stated otherwise. Hplc was carried out with a Waters model 6000 instrument equipped with a refractive index detector. The analyses were carried out on plates spread with 0.25-mm layers of silica gel G (E. Merck, Darmstadt); components were visualized under uv or by charring with sulfuric acid-dichromate solution. Fractions and compounds were tested against P-388 lymphocytic leukemia in the mouse and Eagles 9KB masopharyngeal carcinoma cell culture according to established protocols (9) under auspices of the National Cancer Institute. In the P-388 system an effective response is obtained when the median survival time of treated animals over control animals (% T/C) is $\geq 130\%$. In the KB cell culture system, results are expressed as that dose which inhibits growth to 50% of control growth (ED₅₀). An ED₅₀ of 4 µg/ml is considered effective for pure compounds and 20 µg/ml for crude plant fractions.

SESBANINE (1).—Crude 1, as obtained by hplc (50 mg) was recrystallized from methanol, mp 240–243°; $[\alpha]^{23}D+14.6°$ (c 0.56, MeOH); ir 3510, 3490, 1710, 1690, 1600 cm⁻¹ (KBr); ¹H nmr (CDCl₃-CD₃OD); δ 9.26 (d, 1 Hz, H-8), 8.88 (dd, 6 and 1 Hz, H-6), 7.94 (d, 6 Hz, H-5); the H-10 proton appeared as a multiplet at 4.74 and remaining protons appeared in poorly resolved methods are unkinker 130° error (DMSO 460° 1377 9 (C 1)) 1620 (C 1) 1620 (C 1) H-10 proton appeared as a multiplet at 4.74 and remaining protons appeared in poorty resolved upfield multiplets; ¹³C nmr (DMSO-d6); δ 177.2 (C-1), 163.8 (C-3), 52.1 (C-4), 155.8 (C-4a), 121.7 (C-5), 148.3 (C-6), 153.9 (C-8), 119.6 (C-8a),³ 48.5 (C-9), 72.7 (C-10), 36.1 (C-11), 42.4 (C-12): ¹³C nmr (pyridine-d₅); δ 178.5 (s, C-1), 165.0 (s, C-3), 53.2 (s, C-4), 156.6 (s, C-4a), 154.5 (d, C-8), 49.7 (t, C-9), 73.9 (d, C-10), 37.8 (t, C-11), 43.9 (t, C-12); ms, *m/e* (relative intensity): 232 (12), 214 (6), 204 (3), 199 (4), 175 (100), 162 (10), 157 (13), 129 (11). CI ms, found: M⁺+1, C₁₂H₁₃N₂O₃, *m/e* 233.0907; C₁₂H₁₃N₂O₃ requires 233.0888.

DRUMMONDOL (2).—Compound 2 was isolated as a colorless oil, 145 mg; $[\alpha]^{23}D = -21^{\circ}$ (c 1.03, MeOH); ir (CHCl₃): 3700, 3500, 1725 cm⁻¹; 'H and ¹³C nmr (see table 2); ms, m/e (relative intensity): 222 (3), 204 (8), 164 (7), 136 (12), 125 (17), 123 (12), 99 (34), 82 (45), 71 (33), 69 (25), 55 (52), 43 (100).

DRUMMONDOL p-BROMOBENZOATE (2a).-A 36 mg portion of 2 was dissolved in pyridine (3 ml), 90 mg of p-bromobenzoyl chloride was added, and the resulting solution was allowed to stand overnight at 25°. Pyridine was removed (N_2) , and the residue was partitioned between chloroform and water. Preparative the of the chloroform soluble material (silica, chloroform-Chloroform and water. Freparative tic of the chloroform soluble material (silica, chloroform-methanol 97.5:2.5) then gave **2a** (20 mg) as an amorphous solid; ir (CHCl₃): 3630, 1720, 1600 em⁻¹; ¹H nmr (CDCl₃): δ 0.94 (s, 3H), 1.17 (s, 3H), 1.48 (d, 6Hz, 3H), 2.32 (d, 18Hz, 1H), 2.54 (s, 2H), 2.57 (dd, 18 and 2Hz, 1H), 3.68 (d, 8Hz, 1H), 3.86 (dd, 8 and 2Hz, 1H), 5.58 (m, 6Hz, 1H), 5.92 (d, 15.5Hz, 1H), 6.19 (q, 15.5 and 6Hz, 1H), 7.4-7.9 (m, 4H); ms, *m/e* (relative in-tensity): 222 (21), 204 (15), 185 (40), 183 (42), 164 (21), 136 (40), 125 (66), 123 (82), 82 (100). Found: M⁺-C₇H₅O₂Br, *m/e* 222.1266; C₁₃H₁₈O₃ required 222.1255.

ACKNOWLEDGMENTS

We thank E. C. Baker for assistance with pilot plant extraction, Dr. D. Weisleder and L. Tjarks for nmr spectra, D. King and Dr. W. K. Rohwedder for mass spectra, Dr. G. J. Jordan, Lederle Laboratories, Pearl River, New York, for an FTIR spectrum of sesbanine, and Dr. R. E. Perdue for supplying S. drummondii seed.

Received 1 August 1980

LITERATURE CITED

- J. M. Kingsbury, "Poisonous Plants of the United States and Canada," Prentice-Hall, Englewood Cliffs, New Jersey, 1964, p. 356. R. G. Powell, C. R. Smith, Jr. and R. V. Madrigal, *Planta Med.*, **30**, 1 (1976). R. G. Powell, C. R. Smith, Jr., D. Weisleder, D. A. Muthard and J. Clardy, *J. Am. Chem.* 1.
- 2
- 3. Soc., 101, 2784 (1979). A. S. Kende and T. P. DeMuth, Tetrahedron Lett., 21, 715 (1980).
- 4.
- 5.
- 6.
- 7.
- 8.
- J. C. Bottaro and G. A. Berchtold, J. Org. Chem., 45, 113 (1980).
 J. C. Bottaro and G. A. Berchtold, J. Org. Chem., 45, 1176 (1980).
 B. V. Milborrow, Phytochemistry, 14, 1045 (1975).
 D. S. Bhakumi, P. P. Joshi, H. Uprety and R. S. Kapil, Phytochemistry, 13, 2541 (1974).
 J. A. D. Zeevaart and B. V. Milborrow, Phytochemistry, 15, 493 (1976).
 R. I. Geran, N. H. Greenberg, M. N. MacDonald, A. M. Schumacher and B. J. Abbott, Cancer Chemother. Rep., Part 3, 3, 1 (1972). Q.

³This signal was erroneously quoted (3) as overlapping with the C-5 signal (δ 121.7). Assignments for C-5 and C-8a may be reversed.